# Data Mining - Data

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Data Mining - Data

- 1. Introduction
- 2. Data preprocessing
- 3. CPA with R
- 4. Exercise and home work









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#### Data Mining - Data

## **Definition 1**

- preprocessing of data (normalization, standardization)
- sampling to train and test classifiers
- feature selection with PCA



## **Raw data collection**

datasets are not always in good shape

- different sources of information (Database, Excel, ...)
- missing data, errors

a first pre-processing step is needed

- for example scaling, translation, and rotation of images
- rearrange, remove rows with empty data
- or imputate values, i.e. replace missing values using certain statistics (mean, most common, ...)







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## Normalization

Normalization and other feature scaling techniques are often mandatory in order to make comparisons between different attributes if the attributes were **measured on different scales** (e.g., temperatures in Kelvin and Celsius);

the term normalization is often used synonymous to Min-Max scaling

The scaling of attributes in a certain range, e.g., 0 to 1:

$$X_{norm} = \frac{X - X_{min}}{X_{max} - X_{min}}$$





## Standardization

Another common approach is **standardization** or *scaling to unit-variance* 

- every sample is subtracted by the attribute's mean  $\mu$  and divided by the standard deviation  $\sigma$
- attributes will have the properties of a standard normal distribution ( $\mu = 0$  and  $\sigma = 1$ )

this is done in order not to have one property which will have more influence than the others





## Sampling

split the dataset into a **training** and a **test** dataset to assess precision of method (classifier)

- training dataset is used to train the model
- test dataset evaluate the performance of the final model
- be aware of overfitting: classifiers that perform well on training data but do not generalize well





## **Cross-Validation**

most useful techniques to evaluate different combinations of

- feature (property) selection (take only intersting features)
- dimensionality reduction of learning algorithms (reduce for efficiency)

multiple flavors of cross-validation, most common is  ${\bf k}\mbox{-fold}$  cross-validation



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## K-fold

the original training dataset is split into k different subsets (the so-called folds)

- one fold is retained as test set
- the other k 1 are used for training the model
- calculate the average error rate (and standard deviation) of the folds

use matrix of statistics to see if some fold is different from others





## Feature Selection and Dimensionality Reduction

the key difference between the terms:

- feature selection: keep the original feature axis
- dimensionality reduction: usually involves a transformation technique

the main goals are:

- noise reduction
- increase computational efficiency by retaining only useful (discriminatory) information

avoid overfitting





## **Feature Selection**

retain only those features that are **meaningful** and help build a *good* classifier by reducing the number of attributes (features, properties)

Linear Discriminant Analysis (LDA): supervised, computes the directions (LD) that maximize the separation between multiple classes

Principal Component Analyses (PCA): unsupervised, ignores class labels, find PC that maximize the variance in a dataset



## LDA

### in a nutshell

- project a feature space (a dataset of *k*-dimensional samples) onto a smaller subspace h (where  $h \le k 1$ )
- while maintaining the class-discriminatory information



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## PCA (Principal Component Analysis

- reduce the dimensionality of a data set consisting of a large number of interrelated variable
- retain as much as possible the variation in the data set
- achieved by transforming to a new set of uncorrelated variables (PC)
- ordered so that the first few retain most of the variation present in all of the original variables





## PCA Algorithm (1/2)

- Take the whole dataset consisting of k-dimensional samples ignoring the class labels
- Compute the k-dimensional mean vector (i.e., the means for every dimension of the whole dataset)
- Compute the scatter matrix (alternatively, the covariance matrix) of the whole data set
- Compute eigenvectors (e<sub>1</sub>, e<sub>2</sub>,..., e<sub>k</sub>) and corresponding eigenvalues (λ<sub>1</sub>, λ<sub>2</sub>,..., λ<sub>k</sub>)





## PCA Algorithm (2/2)

- Sort the eigenvectors by decreasing eigenvalues and choose h eigenvectors with the largest eigenvalues to form a k × h dimensional matrix W (where every column represents an eigenvector)
- Use this  $k \times h$  eigenvector matrix to transform the samples onto the new subspace  $y = W^T \times x$  (where x is a  $k \times 1$ -dimensional vector representing one sample and y is the transformed  $h \times 1$ -dimensional sample in the new subspace





# PCA algorithm (2/2)

## **Types of PCA**

- general: apply algorithm to data (no transformation of data)
- **centered**: substract mean to initial values
- reduced: first center then reduce (if variances are significantly different, divide each variables by its standard deviation)







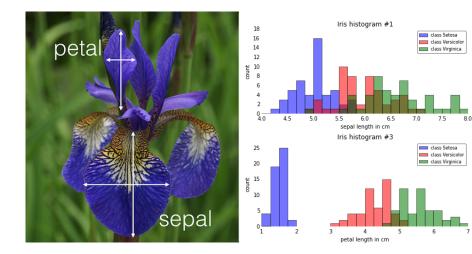
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To have a better understanding of PCA we will use the **IRIS** flowers dataset

- 150 individuals (50 Setosa, 50 Versicolour, 50 Virginica)
- 5 properties:
  - petal length and width
  - sepal length and width (protection for the flower and support for the petals when in bloom)
  - class (Setosa, Versicolour, Virginica)



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#### View the data

#### Dataset already present

| > | head(iris)   |             |              |             |         |
|---|--------------|-------------|--------------|-------------|---------|
|   | Sepal.Length | Sepal.Width | Petal.Length | Petal.Width | Species |
| 1 | 5.1          | 3.5         | 1.4          | 0.2         | setosa  |
| 2 | 4.9          | 3.0         | 1.4          | 0.2         | setosa  |
| 3 | 4.7          | 3.2         | 1.3          | 0.2         | setosa  |
| 4 | 4.6          | 3.1         | 1.5          | 0.2         | setosa  |
| 5 | 5.0          | 3.6         | 1.4          | 0.2         | setosa  |
| 6 | 5.4          | 3.9         | 1.7          | 0.4         | setosa  |



> summary(iris) Sepal.Length Sepal.Width Min. :4.300 Min. 1st Qu.:5.100 Median :5.800 Mean :5.843 3rd Qu.:6.400 Max. :7.900 Species :50 setosa versicolor:50 virginica :50

:2.000 1st Qu.:2.800 Median :3.000 Mean :3.057 3rd Qu.:3.300 Max. :4,400

| Petal.Length |        |  |  |  |
|--------------|--------|--|--|--|
| Min.         | :1.000 |  |  |  |
| 1st Qu.      | :1.600 |  |  |  |
| Median       | :4.350 |  |  |  |
| Mean         | :3.758 |  |  |  |
| 3rd Qu.      | :5.100 |  |  |  |
| Max.         | :6.900 |  |  |  |

| Petal.Width |     |     |     |
|-------------|-----|-----|-----|
| Min.        |     | :0. | 100 |
| 1st         | Qu. | :0. | 300 |
| Medi        | an  | :1. | 300 |
| Mean        |     | :1. | 199 |
| 3rd         | Qu. | :1. | 800 |
| Max.        |     | :2. | 500 |



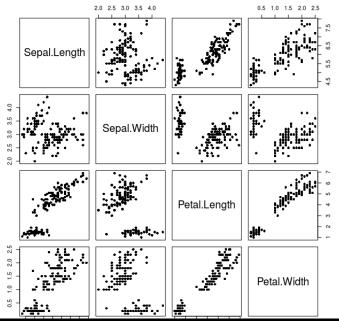
Plot the data to have a better understanding (with hundreds of attribute this is not feasible)

```
> attach(iris)
> plot(iris[,1:4], pch=16)
```

what can we really see ?







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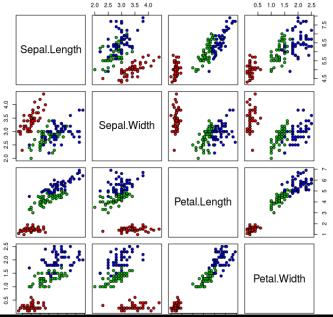
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Plot in function of classes:

- > attach(iris)
- > pairs(iris[1:4],pch = 21, bg = c("red", "green3", "blue")[unclass(iris\$Species)])









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#### In this case

- if you compare Sepal.Length to Sepal.Width:
  - the red dots are separated from the blue and green dots
  - but it will be difficult to differentiate blue from green
- if you compare Petal.Length to Petal.Width the differentiation is simple





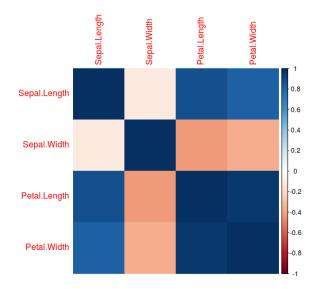
Correlation matrix values

| > cor(iris[,1:4]) |              |             |              |             |  |
|-------------------|--------------|-------------|--------------|-------------|--|
|                   | Sepal.Length | Sepal.Width | Petal.Length | Petal.Width |  |
| Sepal.Length      | 1.0000000    | -0.1175698  | 0.8717538    | 0.8179411   |  |
| Sepal.Width       | -0.1175698   | 1.0000000   | -0.4284401   | -0.3661259  |  |
| Petal.Length      | 0.8717538    | -0.4284401  | 1.0000000    | 0.9628654   |  |
| Petal.Width       | 0.8179411    | -0.3661259  | 0.9628654    | 1.0000000   |  |

Then plot the correlation matrix

```
library(corrplot)
corrplot(cor(iris[,1:4]), method="color")
```







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## **Correlation explanation**

if the correlation value is

- close to 1, then both vectors of values are strongly correlated
- close to 0, then they are not correlated

In this example *Sepal.Length* and *Petal.Length* are strongly correlated, so it is interesting to remove one of them



## What we want to know

- is there a difference between the species ?
- which properties can explain thoses differences ?





## Perform the PCA with R

pca = prcomp(iris[,1:4])

The principal component are obtained with pca\$rotation



### **Principal components**

#### > pca\$rotation

|              | PC1         | PC2         | PC3         | PC4        |
|--------------|-------------|-------------|-------------|------------|
| Sepal.Length | 0.36138659  | -0.65658877 | 0.58202985  | 0.3154872  |
| Sepal.Width  | -0.08452251 | -0.73016143 | -0.59791083 | -0.3197231 |
| Petal.Length | 0.85667061  | 0.17337266  | -0.07623608 | -0.4798390 |
| Petal.Width  | 0.35828920  | 0.07548102  | -0.54583143 | 0.7536574  |

|                 | ( | 0.86 | $\times$ | Petal.Length |
|-----------------|---|------|----------|--------------|
| $PC1 = \langle$ | + | 0.36 | ×        | Sepal.Length |
| $PCI = \langle$ | + | 0.36 | ×        | Petal.Width  |
|                 | _ |      |          | Sepal.Width  |



## Principal components information

pca\$sdev gives the pourcentage of information in each component:

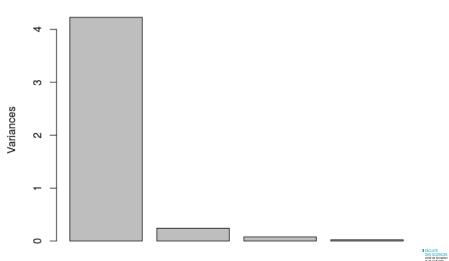
```
> pca$sdev
[1] 2.0562689 0.4926162 0.2796596 0.1543862
> 100 * pca$sdev^2 / sum(pca$sdev^2)
[1] 92.4618723 5.3066483 1.7102610 0.5212184
> sum(100 * (pca$sdev^2)[1:1] / sum(pca$sdev^2))
[1] 92.46187
> sum(100 * (pca$sdev^2)[1:2] / sum(pca$sdev^2))
[1] 97.76852
> sum(100 * (pca$sdev^2)[1:3] / sum(pca$sdev^2))
[1] 99.47878
> pca$rotation
```

Already 92% of the information is given by the first component !

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## Print individuals in function of PC

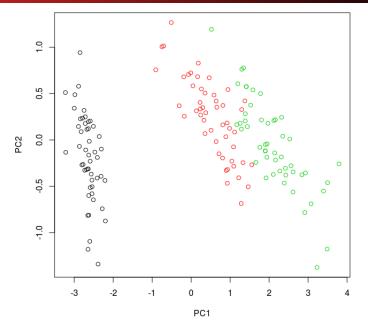
```
> plot(pca$x[,1], rep(0,nrow(iris)), col=iris[,5])
```

#### or

```
> library(ggfortify)
> df <- iris[c(1, 2, 3, 4)]
> autoplot(prcomp(df), data = iris, colour = 'Species')
> autoplot(prcomp(df), data = iris, colour = 'Species',
    label = TRUE, label.size = 3)
```



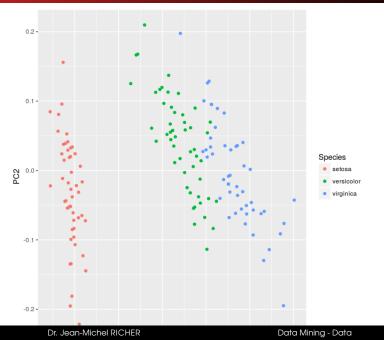
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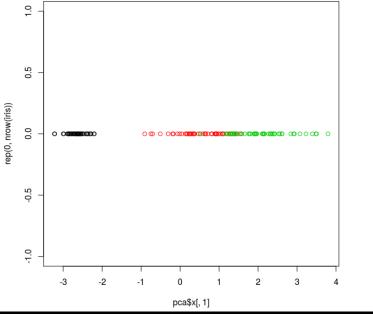
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## **Final interpretation**

- setosa (red) can be identified clearly
- as the PC 1 is composed of 0.86 × Petal.Length, it is the length of the petal that can help differentiate species



## More information

## Follow the following links

- http://www.sthda.com/english/wiki/print.php?id=206
- https://www.analyticsvidhya.com/blog/2016/03/ practical-guide-principal-component-analysis-python/







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## PCA with python

install the sklearn package and try to do as R

- display information: data, correlation matrix
- perform the PCA, print results
- try to plot data





## PCA with Weka

- write a report explaining how to perform PCA using Weka
- give detail of each step







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